

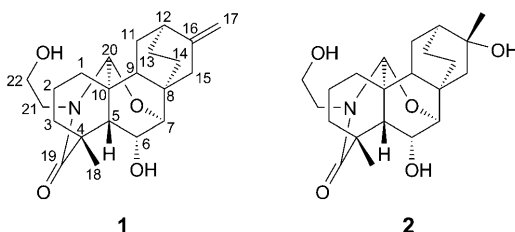
Two New Diterpenoid Lactams from *Spiraea japonica* var. *ovalifolia*

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Two new diterpenoid alkaloids, spiramilactams A (**1**) and B (**2**), were isolated from the basic fraction of a MeOH extract of whole plants of *Spiraea japonica* var. *ovalifolia*. Their structures were established on the basis of extensive spectroscopic and mass-spectrometric analyses.

Introduction. – The *Spiraea japonica* complex (Rosaceae) was shown to contain diterpenoid alkaloids and diterpenoids, containing 22 hetisine-type alkaloids, 37 atisine-type alkaloids, and eight atisane-type diterpenoids [1–3]. Among them, some atisine-type alkaloids displayed significant bioactivity concerning anti-inflammation, antiplatelet aggregation, and neuroprotective effects [4–7]. In our ongoing search for additional diterpenoid alkaloids from the above plant complex, a reinvestigation on the chemical constituents of *Spiraea japonica* var. *ovalifolia* collected in Songming County of Yunnan Province, P. R. China, led to the isolation of six compounds including two novel atisine-type diterpenoid lactams, spiramilactams A (**1**) and B (**2**), along with four known diterpenoid alkaloids, namely spiramines A–D. This is the first report of the diterpenoid alkaloids from the *S. japonica* complex with α -configuration for the HO–C(6) group. Details of the isolation and structural elucidation of the two new diterpenoid lactams are presented below.



Results and Discussion. – Spiramilactam A (**1**) was obtained as an optically active, white powder. The molecular formula was established as $C_{22}H_{31}NO_4$ by the HR-ESI-MS (m/z 374.2326 ($[M + H]^+$; calc. 374.2331)). The IR spectrum indicated the presence of OH groups (3420 cm^{-1}), a CO group (1735 cm^{-1}), and a C=C bond (1632 cm^{-1}). The $^1\text{H-NMR}$ spectrum of **1** (Table) showed signals for one tertiary Me group at $\delta(\text{H})$ 1.53 (s, Me(18)), one O-bearing CH_2 group at $\delta(\text{H})$ 4.12 (t, $\text{CH}_2(22)$), an exo- CH_2 group at $\delta(\text{H})$ 4.80 and 4.94 (2 br. s, $\text{CH}_2(17)$), and two O-bearing CH groups

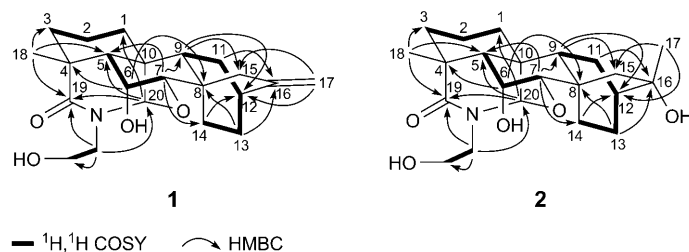
at $\delta(\text{H})$ 3.75 (*d*, H–C(7)) and 4.46 (*dd*, H–C(6)). The ^{13}C -NMR and DEPT spectrum of **1** (Table) indicated 22 C-atom signals, consisting of one Me group, nine CH_2 groups, including one CH_2O group, six CH groups, including two O–CH groups, four quaternary C-atoms, including a CO group at $\delta(\text{C})$ 175.4, and two olefinic resonances at $\delta(\text{C})$ 108.0 (*t*) and 152.0 (*s*). The above-mentioned ^1H - and ^{13}C -NMR data of **1** revealed that compound **1** was an atisine-type lactam derivative [8–11]. Comparison of the ^1H - and ^{13}C -NMR spectral data of **1** with those of spiramine Y [8] showed that the two compounds possessed similar C-atom skeletons, except for the absence of an AcO group in the former compound. The two O-bearing CH groups at $\delta(\text{C})$ 71.3 (C(6)) and 74.7 (C(7)) correlated with CH groups at $\delta(\text{H})$ 4.46 (H–C(6)) and 3.75 (H–C(7)), respectively, in the HMQC experiments, and the two CH groups showed cross peaks in the ^1H , ^1H -COSY spectra, suggesting that their H-atoms are vicinal to each other. The correlations of H–C(7) with C(5), C(9), C(14), C(15), and C(20) in the HMBC spectrum confirmed that C(7) was connected to C(20) through an O-bridge. HMBC correlations of H–C(20) with C(5), C(19), and C(21) suggested that C(19), C(20), and C(21) were connected to each other through a N-atom. Further-

Table. ^1H - and ^{13}C -NMR Data of Compounds **1** and **2**. In $\text{C}_5\text{D}_5\text{N}$; δ in ppm, *J* in Hz.

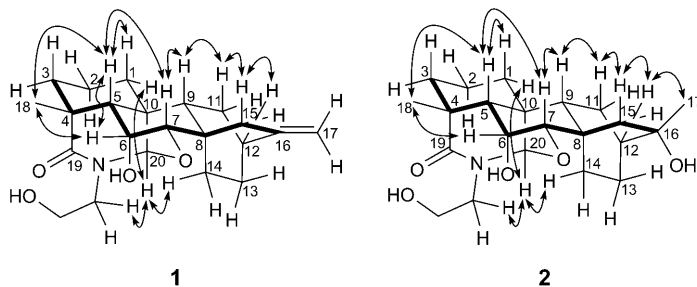
	1 ^{a)}		2 ^{b)}	
	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$
$\text{CH}_2(1)$	29.2 (<i>t</i>)	1.78 (<i>dd</i> , <i>J</i> = 5.0, 14.5), 1.20 (<i>ddd</i> , <i>J</i> = 5.0, 6.5, 14.5)	29.1 (<i>t</i>)	1.89–1.86 (<i>m</i>), 1.37–1.35 (<i>m</i>)
$\text{CH}_2(2)$	25.9 (<i>t</i>)	1.65–1.63 (<i>m</i>), 1.49–1.47 (<i>m</i>)	23.9 (<i>t</i>)	1.58–1.55 (<i>m</i>), 1.53–1.51 (<i>m</i>)
$\text{CH}_2(3)$	20.5 (<i>t</i>)	1.93–1.89 (<i>m</i>), 1.46–1.42 (<i>m</i>)	20.6 (<i>t</i>)	1.92–1.90 (<i>m</i>), 1.43–1.40 (<i>m</i>)
C(4)	44.2 (<i>s</i>)	–	44.3 (<i>s</i>)	–
H–C(5)	58.8 (<i>d</i>)	1.60–1.58 (<i>m</i>)	59.4 (<i>d</i>)	1.76 (<i>br. s</i>)
H–C(6)	71.3 (<i>d</i>)	4.46 (<i>dd</i> , <i>J</i> = 2.8, 4.4)	71.7 (<i>d</i>)	4.50 (<i>dd</i> , <i>J</i> = 1.2, 3.0)
H–C(7)	74.7 (<i>d</i>)	3.75 (<i>d</i> , <i>J</i> = 4.4)	75.2 (<i>d</i>)	3.78–3.76 (<i>m</i>)
C(8)	37.0 (<i>s</i>)	–	36.9 (<i>s</i>)	–
H–C(9)	46.7 (<i>d</i>)	1.70 (<i>dd</i> , <i>J</i> = 4.0, 8.5)	42.5 (<i>d</i>)	1.31–1.29 (<i>m</i>)
C(10)	34.6 (<i>s</i>)	–	34.8 (<i>s</i>)	–
$\text{CH}_2(11)$	40.0 (<i>t</i>)	1.88–1.86 (<i>m</i>), 1.40–1.37 (<i>m</i>)	40.1 (<i>t</i>)	1.84–1.81 (<i>m</i>), 1.32–1.30 (<i>m</i>)
H–C(12)	37.2 (<i>d</i>)	2.30 (<i>t</i> , <i>J</i> = 4.5)	39.0 (<i>d</i>)	1.82–1.80 (<i>m</i>)
$\text{CH}_2(13)$	26.9 (<i>t</i>)	1.61–1.59 (<i>m</i>), 1.33–1.31 (<i>m</i>)	23.7 (<i>t</i>)	2.18–2.17 (<i>m</i>), 1.32–1.30 (<i>m</i>)
$\text{CH}_2(14)$	27.4 (<i>t</i>)	2.08 (<i>dd</i> , <i>J</i> = 2.0, 11.0), 1.38–1.35 (<i>m</i>)	27.6 (<i>t</i>)	2.08–2.06 (<i>m</i>), 1.26–1.24 (<i>m</i>)
$\text{CH}_2(15)$	40.1 (<i>t</i>)	3.37 (<i>d</i> , <i>J</i> = 15.0), 2.22 (<i>d</i> , <i>J</i> = 15.0)	47.6 (<i>t</i>)	1.44–1.46 (<i>m</i>)
C(16)	152.0 (<i>s</i>)	–	72.7 (<i>s</i>)	–
$\text{CH}_2(17)$ / Me(17)	108.0 (<i>t</i>)	4.94 (<i>br. s</i>), 4.80 (<i>br. s</i>)	30.5 (<i>q</i>)	1.47 (<i>s</i>)
Me(18)	21.6 (<i>q</i>)	1.53 (<i>s</i>)	21.7 (<i>q</i>)	1.53 (<i>s</i>)
C(19)	175.4 (<i>s</i>)	–	175.8 (<i>s</i>)	–
H–C(20)	86.2 (<i>d</i>)	5.24 (<i>d</i> , <i>J</i> = 1.6)	86.7 (<i>d</i>)	5.30 (<i>d</i> , <i>J</i> = 1.7)
$\text{CH}_2(21)$	50.1 (<i>t</i>)	4.32 (<i>ddd</i> , <i>J</i> = 2.0, 6.0, 13.5), 3.71 (<i>ddd</i> , <i>J</i> = 2.0, 6.0, 13.5)	50.2 (<i>t</i>)	4.28 (<i>ddd</i> , <i>J</i> = 2.0, 5.6, 11.2), 3.75–3.73 (<i>m</i>)
$\text{CH}_2(22)$	60.9 (<i>t</i>)	4.12 (<i>t</i> , <i>J</i> = 6.0)	60.9 (<i>t</i>)	4.13 (<i>t</i> , <i>J</i> = 6.2)

^{a)} Recorded at 500 MHz. ^{b)} Recorded at 400 MHz.

more, analysis of the $^1\text{H},^1\text{H}$ -COSY spectra of **1** established three other fragments: $\text{CH}_2(1)\text{--CH}_2(2)\text{--CH}_2(3)$, $\text{CH}(9)\text{--CH}_2(11)\text{--CH}(12)\text{--CH}_2(13)\text{--CH}_2(14)$, and $\text{CH}_2(21)\text{--CH}_2(22)$, as shown with bold bonds in *Fig. 1*. The location of the exo- CH_2 group at C(16) was supported by the correlations between $\text{CH}_2(17)$ ($\delta(\text{H})$ 4.80, 4.94) and C(12), C(15), and C(16), while the presence of a CO group at C(19) was confirmed by the correlations of $\text{CH}_2(3)$, H–C(5), Me(18), H–C(20), and $\text{CH}_2(21)$ with the CO group ($\delta(\text{C})$ 175.4). The relative configuration of **1** was elucidated by the help of a ROESY spectrum as shown in *Fig. 2*. The ROESY correlations of H–C(5) with H–C(6), H–C(7), and Me(18) implied the α -configuration for the 6-OH group and β -configuration for H–C(7). Thus, the structure of spiramilactam A (**1**) was elucidated to be as shown in **1**.



*Fig. 1. Key $^1\text{H},^1\text{H}$ -COSY and HMBC correlations of compounds **1** and **2***



*Fig. 2. Significant ROESY correlations within compounds **1** and **2***

Spiramilactam B (**2**), obtained as a colorless gum, had the molecular formula of $\text{C}_{22}\text{H}_{33}\text{NO}_5$ as determined by a *pseudo*-molecular ion peak in the HR-ESI-MS (392.2440 ($[M + \text{H}]^+$; calc. 392.2436)). Step-by-step comparison of the ^1H - and ^{13}C -NMR spectral data of **2** with those of **1** (*Table*) revealed that most signals of **2** were similar to those of spiramilactam A (**1**), except for the disappearance of an exocyclic $\text{C}=\text{C}$ bond and the presence of an O-bearing quaternary C-atom ($\delta(\text{C})$ 72.7, C(16)) and a tertiary Me group ($\delta(\text{H})$ 1.47) in compound **2**. Therefore, it was supposed that **2** was derived from **1** by hydroxylation at the exocyclic $\text{C}=\text{C}$ bond, which was confirmed by the mass difference of $\Delta m/z = 18$ and the HMBC spectrum. In the HMBC spectrum of **2**, correlations of Me(17) ($\delta(\text{H})$ 1.47) with C(12), C(15), and C(16) were observed (*Fig. 1*). The relative configuration of the OH group attached at C(16) was determined

by a ROESY experiment (Fig. 2). The Me(17) signal showed ROESY correlations with H–C(12), suggesting β -orientation for Me(17). Therefore, the structure of spiramylactam B (**2**) was determined as shown in **2**.

Experimental Part

General. MPLC: Büchi Pump Module C-605, Büchi Pump Manager C-615, and Büchi Fraction Collector C-660. Column chromatography (CC): silica gel (SiO₂; 200–300 mesh; silica gel H, Qingdao Marine Chemical Ltd.Co.), or Sephadex LH-20 (Pharmacia). TLC: silica-gel plates; visualization by spraying with Dragendorff's reagent. Optical rotations: JASCO DIP-370 digital polarimeter. IR Spectra: Bio-Rad FTS-135 spectrometer, KBr pellets; in cm⁻¹. NMR Spectra: Bruker AM-400 instrument (¹H: 400, ¹³C: 100 MHz) or Bruker DRX-500 instrument (¹H: 500, ¹³C: 125 MHz); δ in ppm rel. to TMS as internal standard, J in Hz. EI-MS: VG Auto Spec-3000 mass spectrometer; in m/z . HR-ESI-MS: API Qstar Pulsar LC/TOF instrument.

Plant Material. The whole plants of *S. japonica* var. *ovalifolia* were collected in July 2007 from Songming, Yunnan Province, P. R. China, and identified by Dr. Zhao-Yang Zhang of Kunming Botanical Garden. A voucher specimen was deposited at the State Key Laboratory of Phytochemistry and Plant Resources in West China, Chinese Academy of Sciences.

Extraction and Isolation. The air-dried whole plants of *S. japonica* var. *ovalifolia* (20 kg) were grounded and extracted with MeOH under reflux (3 \times 50 l, each 3 h). The extracts were condensed *in vacuo* to afford a crude mixture which was dissolved in 3% HCl (5 l) soln. and filtered. The acidic soln. was basified with 5% aq. NaOH to pH 11 and then extracted with CHCl₃. Evaporation of CHCl₃ gave a crude alkaloid mixture (130 g) which was subjected to CC (SiO₂; petroleum ether (PE)/AcOEt/Et₃N 40:10:1 to 10:10:1) to five fractions (*Fr. I–V*). Repeated separation of *Fr. I* and *II* over SiO₂ by MPLC (PE/AcOEt/Et₃N 80:2:1 to 8:2:1) gave spiramines A (200 mg), B (150 mg), C (140 mg), and D (110 mg), and a mixture of spiramines A and B (4.5 g), as well as a mixture of spiramines C and D (2.3 g). *Fr. V* (3.0 g) was purified by repeated MPLC (silica gel H, PE/AcOEt/Et₃N 4:1:0.2 to 1:1:0.2) and Sephadex LH-20 (CHCl₃/MeOH 1:1) to yield **1** (40 mg) and **2** (21 mg).

Spiramylactam A (= (6 α ,7 α)-6-Hydroxy-21-(2-hydroxyethyl)-4-methyl-7,20-epoxyatid-16-en-19-one; **1**). White powder. $[\alpha]_D^{25} = -83.3$ ($c = 0.57$, MeOH). IR (KBr): 3420, 2930, 2871, 1735, 1632, 1470, 1316, 1063, 1011, 885. ¹H- and ¹³C-NMR: Table. FAB-MS (pos.): 374 (100, $[M + H]^+$). HR-ESI-MS: 374.2326 ($[M + H]^+$, C₂₂H₃₂NO₄⁺; calc. 374.2326).

Spiramylactam B (= (6 α ,7 α)-6,16-Dihydroxy-21-(2-hydroxyethyl)-4-methyl-7,20-epoxyatidan-19-one; **2**). Colorless gum. $[\alpha]_D^{25} = -53.2$ ($c = 0.85$, MeOH). IR (film): 3406, 2985, 2878, 1593, 1474, 1346, 1066, 1011, 807, 765. ¹H- and ¹³C-NMR: Table. EI-MS: 391 (25, M^+), 373 (29, $[M - H_2O]^+$), 363 (100), 348 (83), 320 (33), 256 (38), 243 (88), 215 (36), 196 (95), 157 (35), 149 (55), 95 (57), 69 (66). HR-ESI-MS: 392.2440 ($[M + H]^+$, C₂₂H₃₄NO₅⁺; calc. 392.2431).

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